Dose-dependent vascular response following delivery of sirolimus via fast releasing, biodegradable polymer stent matrix: an experimental study in the porcine coronary model of restenosis

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Abstract

Background: Fast releasing, rapamycin-eluting stents, although safe, showed inferior results with regard to inhibition of restenosis.

Aim: Therefore, we report vascular effects of a novel, biodegradable polymer stent matrix with elevated sirolimus dose and fast release kinetics (ed-frSES, Alex, Balton) in the porcine coronary in-stent restenosis model.

Methods: A total of 19 stents were implanted with 120% overstretch in the coronary arteries of seven domestic pigs: seven ed-frSES with 1.3 μg/mm² of sirolimus, eight frSES with 1 μg/mm² of sirolimus, and eight bare metal stents (BMS). For the following 28 days, coronary angiography was performed, animals were sacrificed, and the stented segments harvested for histopathological evaluation.

Results: In angiography at 28 days the late lumen loss was lowest in the elevated dose sirolimus eluting stent (SES) (ed-frSES: 0.20 ± 0.2 vs. frSES: 0.80 ± 0.5 vs. BMS: 0.96 ± 0.5 mm, p < 0.01). This was confirmed in the morphometric evaluation in histopathology as represented by a significant and dose-dependent decrease in the percentage area of stenosis (ed-frSES: 22.4 ± 12.7% vs. frSES: 35 ± 10.7% vs. BMS: 47.5 ± 12.5%, p < 0.01). There was no peri-strut inflammation in any of the groups. However, the endothelialisation score was numerically not meaningfully decreased in ed-frSES (ed-frSES: 2.93 vs. frSES: 3 vs. BMS: 3, p = 0.05). Signs of fibrin were also noted in ed-frSES (ed-frSES: 0.4 vs. frSES: 0 vs. BMS: 0, p = 0.05).

Conclusions: Sirolimus dose-dependent vascular response was reported. The elevated dose, fast releasing SES shows satisfactory vascular healing, similar to regular dose, fast release SES, with improved efficacy in restenosis inhibition.

Key words: sirolimus eluting stent, fast release, biodegradable polymer, porcine coronary model

INTRODUCTION

The balance between vascular healing process and sustained efficacy following percutaneous coronary intervention has been a challenge for nearly two decades. First-generation drug eluting stents (DES), although efficacious in restenosis inhibition when compared to bare metal stents (BMS), caused risk of late stent thrombosis due to increased inflammation, excessive fibrin deposition, and, as a consequence, impaired neointimal and endothelial coverage [1, 2]. Second-generation DES, with the utilisation of novel anti-proliferative and inflammatory rapamycin analogues along with biocompatible or biodegradable polymers, improved significantly the clinical
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Additionally, it has been shown that the optimisation of drug release kinetics may influence the vascular response and efficacy. Early generation zotarolimus eluting stent (ZES), with fast release kinetics provided superior safety, with very low risk of stent thrombosis; however, the efficacy in restenosis inhibition was inferior when compared to slow-release ZES [7]. In our recent experiments we tested the pharmacokinetics of a fast-release, regular dose (1 μg/mm²) sirolimus and biodegradable polymer cobalt chromium stent (FR-SES), which has shown 95% of drug elution within 90 days in an in-vivo setting and complete healing at 30 days [8]; however, the efficacy was limited, similarly to previously cited reports on ZES. In the current study we test the hypothesis that sirolimus dose increase, as well as release kinetics, may influence the vascular healing response. Therefore, the dose of sirolimus on an identical biodegradable-polymer stent matrix was elevated by 30% and vascular response evaluated in the coronary in-stent restenosis model of a swine.

**METHODS**

**Device description**

All stents used in this study utilised the L605 cobalt chromium alloy platform with a strut thickness of 70 μm and closed cell design, which served as a bare metal control (Coflexus, Batlon). Sirolimus eluting stents (SES) are covered with a fully biodegradable multilayer structure containing a copolymer of poly-lactic and glycolic acid and sirolimus. The total mass of the polymer on a 3.0 × 15 mm stent does not exceed 360 μg. Experimental studies in the porcine in-stent restenosis model at eight weeks showed nearly full polymer biodegradation and 95% drug release of initial drug load (Fig. 1) [8]. The studied, elevated dose SES (ed-frSES, Alex, Batlon, Poland) employs sirolimus at 1.3 μg/mm², whereas the reference stent was at a regular dose of 1 μg/mm² (frSES, regular dose, fast releasing sirolimus eluting stent). All stents were 3.0 mm and 3.5 mm in diameter and 15 mm in length.

![Figure 1. In vivo sirolimus tissue retention expressed as percentage of drug load on a stent depicting fast-release profile](image)

**Study design**

A study flow chart is presented in Figure 2. A total of seven domestic swine of both genders were included. All animals ranged from five to seven months of age with an average weight of around 45 kg at the time of enrolment. Middle arterial segments without side branches of all three coronary arteries (RCA, LAD, LCX) were screened. After live quantitative coronary analysis (QCA) evaluation of 21 segments 19 segments were eligible to ensure 120% overstretch for stent implantation and inclusion. Following randomisation of vessel segments in a 2:2:1 fashion, a total of eight elevated dose, fast-release sirolimus eluting stent (ed-frSES; study group), seven frSES (reference group), and four BMS controls were implanted with a suitable pressure required for anticipated overstretch diameter. Additional inflations were performed based upon the target site diameter. The animals were followed up for 28 days. Subsequently, control coronary angiography was performed and the swine were sacrificed. All arterial segments were dissected and harvested for pathological and histomorphometric analysis. All interventions and analyses were blinded to operators and investigators.

**Experimental procedures**

The study protocol was approved by the Local Ethics Committee for animal research. All animals received a standard of care outlined in the study protocol and in accordance with the act of animal welfare and the “Principles of Care of Laboratory Animals” [9].

Three days prior to the procedure dual antiplatelet therapy consisting of 75 mg of clopidogrel and aspirin per day was initiated and continued until the termination. All pigs were fasted overnight before stent implant procedure. Animals were pre-mediated with atropine (0.5 mg) and subsequently sedated with intramuscular ketamine hydrochloride (20 mg/kg) and xylazine (2 mg/kg), intubated, and anesthetised with an intravenous propofol bolus (20–40 mg) followed by a continuous infusion (2–4 mg/kg/h). Electrocardiogram and blood pressure were continuously monitored. A vascular sheath (6 F) was placed in the right or left femoral artery utilising the Seldinger technique. Anticoagulation with heparin was achieved (3,000–10,000 U) to maintain a coagulation time ≥ 250 s. Following coronary angiography all coronary vessels were sized for proper stent implantation after live QCA analysis.

All pigs were anaesthetised and prepared in the same fashion as described above at 28 days following stent implantation, to perform control coronary angiography, and subsequently humanely sacrificed with pentobarbital overdose.

**Quantitative coronary analysis**

Coronary arteries angiographies were obtained using a Siemens Coroskop Millenium Edition angiographic unit. A Judkins Right, 6 French guiding catheter was utilised to obtain coro-
nary angiography and stent implantation. QCA analysis was performed in a blinded fashion using QAngio XA Software version 7.1.14.0 (Medis Medical Imaging Systems) from two contralateral projections. The baseline and 28-day follow-up reference vessel diameters (RVD) were taken from the proximal and distal portion of the treated segments using the guiding catheter as a standard for measurement. The balloon-to-artery ratio was calculated. The percentage diameter stenosis (%DS) at follow-up was calculated as: \([1–(\text{MLD/RVD})] \times 100\%\), where MLD is the minimal lumen diameter.

**Histological analysis**

Following vessel harvesting, stented segments were immersed in normal buffered formalin 10%. For light microscopy all treated vessels were embedded in methyl methacrylate, and then 40–50 microns sections from the proximal, mid, and distal portion of each stented segment were obtained. These sections were stained with haematoxylin and eosin (H&E). The cross-sectional areas (external elastic lamina [EEL], internal elastic lamina [IEL] and lumen area) of each section were measured. Neointimal thickness was measured as the distance from the inner surface of the stent struts to the luminal border. The following measures were used to calculate vessel layer areas: Media = EEL – IEL; Neointima = IEL – lumen area; % area stenosis = [1 – (lumen area / IEL area)] \(\times\) 100. All sections were evaluated using semi-quantitative scoring criteria. To evaluate the amount of injury, criteria defined by Schwartz et al. [10] were used: 0 = IEL intact, 1 = IEL lacerated, 2 = media lacerated, and 3 = EEL lacerated. To evaluate the extent of peri-strut inflammatory reaction, the following grading system by Kornowski et al. [11] was used: 0 = minimal inflammatory response around strut, 1 = few inflammatory cells around strut, 2 = mild to moderate inflammation, can extend into but do not efface surrounding tissue, and 3 = dense and thick peri-strut aggregate of inflammatory cells, effacing surrounding tissue. Each strut in the section was scored and the mean inflammation and injury score for each section was calculated and reported. The adventitial inflammation score is based on the following criteria: 1 = mild, 2 = moderate, and 3 = heavy peripheral inflammatory infiltration. The endothelialisation score was described as the percentage of endothelial coverage of the arterial circumference: 0 = < 25%, 1 = 25–75%, 2 = 76–95%, and 3 = complete. The fibrin deposition was assessed as: 0 — none to focal, 1 — mild involving < 10% of artery circumference, 2 — moderate involving of 10–25% artery circumference, and 3 — heavy, involving > 25% of artery circumference.

**Statistical analysis**

Normally distributed parametric data are expressed as average and standard deviation, and as median and interquartile range in cases of skewed distribution. When equal variance and normality were observed, one-way analysis of variance (ANOVA) with Student-Newman-Keuls post-ANOVA tests were used to test for differences in variables between stent types. When either equal variance test or normality test failed, Kruskal-Wallis test (with Dunn’s method for post-hoc group comparison) was conducted. A value of \(p \leq 0.05\) was considered statistically significant. MedCalc Statistical Software version 14.12.0 (MedCalc Software bvba, Ostend, Belgium; http://www.medcalc.org; 2014) was used for the data analysis.

**RESULTS**

**Quantitative coronary angiography**

A summary of the QCA analysis is presented in Table 1. The vessels included in the study did not differ with regard to sizes as shown by similar RVDs between groups. The applied stent balloon inflation ensured comparable 120–125% overstretch,
acute gain, and vessel diameters after procedure. The vessel flow was normal in all arteries and there were no dissections requiring additional stent implantation.

At 28 days the in-stent minimal lumen diameter in ed-frSES was higher by 20% when compared to SES (p = 0.007) and 10% when compared to BMS (p = 0.18). This corresponded with %DS, which in the ed-frSES was reduced by 53% and 64% when compared to frSES (p = 0.002) and BMS (p = 0.004), respectively. Finally, the late lumen loss was also reduced by 75% and 80% when compared to frSES (p = 0.01) and BMS subsequently (p = 0.001) (Fig. 3).

**Morphometric analysis**

A summary of the morphometric analysis is presented in Table 2. In the pathological, morphometric analysis the vessels sizes were comparable, as shown by similar external elastic lamina areas between groups. Additionally, stent areas were also similar. The neointimal area in the ed-frSES was reduced by nearly 60% (p = 0.004) and 40% (p = 0.082) when compared to BMS and frSES, consecutively. Correspondingly, the percentage area of stenosis was lowest in the ed-frSES, with a reduction of nearly 50% when compared to BMS (p = 0.016) and 35% when compared with frSES (p = 0.05). The area stenosis was also reduced by 27% in the frSES; however, it was not statistically significant (Fig. 3).

The healing and biocompatibility profile of all tested devices is presented in Figure 4. The injury, peri-strut, and adventitial inflammation were modest and comparable between groups. Endothelialisation score was statistically significantly, but marginally lower (by less than 3%) in ed-frSES (ed-frSES: 2.93 vs. frSES: 3 vs. BMS: 3, p = 0.05). Similarly, the fibrin deposition and lipid loading severity were low, but significantly higher in the ed-frSES. The representative stent cross sections are presented in Figure 5.

**DISCUSSION**

In the current study we evaluate the vascular response to different doses of fast releasing sirolimus and biodegradable polymer coated cobalt chromium stents in the porcine, coronary in-stent restenosis model. The elevated dose (1.3 μg/mm²) stent is compared with a regular dose (1 μg/mm²) reference stent, which both release 95% of the drug within three months from a biodegradable polymer platform in vivo [8]. Structurally identical BMS served as a control. The correctness of the methodology was shown by comparable vessel sizes and stents enrolled. Furthermore, the induced degrees of injuries as expressed by the balloon-to-artery ratios were similar between the groups. At one-month follow-up the angiographic measures of neointimal hyperplasia (late lumen loss and percentage diameter stenosis) were four- and five-fold lower when compared to regular dose, fast-release SES and BMS, consecutively (p < 0.01). In the histopathological analysis, the

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**Table 1. Baseline and 28-day follow-up vessel characteristics assessed by qualitative coronary angiography**

<table>
<thead>
<tr>
<th></th>
<th>frSES (n = 8)</th>
<th>ed-frSES (n = 7)</th>
<th>BMS (n = 4)</th>
<th>P (ANOVA)</th>
</tr>
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<tbody>
<tr>
<td><strong>Baseline</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>RVD [mm]</td>
<td>2.54 ± 0.4</td>
<td>2.43 ± 0.2</td>
<td>2.86 ± 0.4</td>
<td>0.22</td>
</tr>
<tr>
<td>Balloon to artery ratio</td>
<td>1.24 ± 0.19</td>
<td>1.27 ± 0.1</td>
<td>1.20 ± 0.1</td>
<td>0.82</td>
</tr>
<tr>
<td>Acute gain</td>
<td>0.31 ± 0.4</td>
<td>0.32 ± 0.17</td>
<td>0.47</td>
<td>0.59</td>
</tr>
<tr>
<td><strong>28 days</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>In MLD [mm]</td>
<td>2.05 ± 0.4</td>
<td>2.56 ± 0.2*</td>
<td>2.36 ± 0.6</td>
<td>0.01</td>
</tr>
<tr>
<td>RVD [mm]</td>
<td>2.50 ± 0.3</td>
<td>2.74 ± 0.2</td>
<td>2.89 ± 0.2</td>
<td>0.06</td>
</tr>
<tr>
<td>%DS</td>
<td>14.83 (12.3–21.04)</td>
<td>6.94* (5.75–8.30)</td>
<td>18.6 (11.81–43.01)</td>
<td>0.01</td>
</tr>
<tr>
<td>LLL [mm]</td>
<td>0.80 ± 0.5</td>
<td>0.20 ± 0.1*#</td>
<td>0.96 ± 0.5</td>
<td>0.008</td>
</tr>
</tbody>
</table>

*p < 0.05 for ed-SES vs. SES; #p < 0.05 for edSES vs. BMS; BMS — bare metal stent; %DS — percentage diameter stenosis; ed-frSES — elevated dose, fast-release sirolimus eluting stent; frSES — regular dose, fast-release sirolimus eluting stent; LLL — late lumen loss; MLD — minimal lumen diameter; RVD — reference vessel diameters; SES — sirolimus eluting stent

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**Figure 3.** Angiographic and histomorphometric parameters representing device efficacy; BMS — bare metal stent; ed-frSES — elevated dose, fast-release sirolimus eluting stent; frSES — regular dose, fast-release sirolimus eluting stent; *p < 0.05 for ed-frSES vs. frSES; #p < 0.05 for ed-frSES vs. BMS.
Table 2. Histopathological results of vessel morphometry, biocompatibility, and healing at 28 days of follow-up

<table>
<thead>
<tr>
<th></th>
<th>fr-SES (n = 8)</th>
<th>ed-frSES (n = 7)</th>
<th>BMS (n = 4)</th>
<th>P (ANOVA)</th>
</tr>
</thead>
<tbody>
<tr>
<td>EEL area [mm²]</td>
<td>10.67 ± 1.8</td>
<td>9.4 ± 1.6</td>
<td>13.25 ± 2.7</td>
<td>0.021</td>
</tr>
<tr>
<td>Stent area [mm²]</td>
<td>7.7 ± 1.1</td>
<td>7.91 ± 1.40</td>
<td>9.36 ± 1.9</td>
<td>0.07</td>
</tr>
<tr>
<td>Lumen area [mm²]</td>
<td>4.49 ± 0.8</td>
<td>6.1 ± 1.7</td>
<td>4.64 ± 0.76</td>
<td>0.069</td>
</tr>
<tr>
<td>Neointimal area [mm²]</td>
<td>2.45 ± 0.9*</td>
<td>1.65 ± 0.7*</td>
<td>4.33 ± 1.7</td>
<td>0.003</td>
</tr>
<tr>
<td>Area of stenosis [%]</td>
<td>35 ± 10.7%</td>
<td>22.4 ± 12.7%*</td>
<td>47.5 ± 12.5%</td>
<td>0.005</td>
</tr>
</tbody>
</table>

*p < 0.05 vs. BMS; BMS — bare metal stent; ed-frSES — elevated dose, fast-release sirolimus eluting stent; frSES — regular dose, fast-release sirolimus eluting stent; EEL — external elastic lamina

Figure 4. Qualitative histopathological analysis representing healing and biocompatibility; BMS — bare metal stent; ed-frSES — elevated dose, fast-release sirolimus eluting stent; frSES — regular dose, fast-release sirolimus eluting stent

Figure 5. Representative histopathological stent-cross section; upper panel: stent-cross section (A. ed-frSES; B. frSES; C. BMS); lower panel: high power: peri-strut magnification 20×; abbreviations as in Figure 4
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As a consequence the biological effect (restenosis inhibition) would not affect the sirolimus receptor binding capabilities, and as a consequence the biological effect was reported. This result contrary to the previously-cited study might be explained by the analytical rather than empirical methodology and multifactorial action of sirolimus [24].

The safety and efficacy of a studied ed-frSES (Alex, Balton) was shown in the first-in-man study [25], in which temporal vascular healing was evaluated with serial Optical Coherence Tomography (OCT) evaluation. The favourable healing profile was confirmed at three months, at which time 97% of struts were covered by neointima. The efficacy was sustained until 12 months with in-stent late-lumen loss of 0.14 mm. Longer follow-up in the clinical setting is required to establish a full safety profile of the studied stent.

Limitations of the study

The limitations of this study include the nature of the experimental preclinical model as a human clinical surrogate and utilisation of a healthy domestic swine, without underlying disease. Furthermore, no intravascular imaging was utilised to prevent injury to endothelium caused by mechanical pullback to provide unbiased histopathological results. Histopathological evaluation was performed with H&E staining only.

CONCLUSIONS

In conclusion, the dose-dependent vascular effect of sirolimus, rapidly eluted from the biodegradable polymer and stent matrix, was observed. The ed-frSES provided efficacious neointimal hyperplasia inhibition when compared to regular dose frSES; however, the healing was satisfactory. These findings support the notion that not only drug release kinetics, but also the dose of the eluted drug influences the biological response.

The clinical implications are important. The combination of fast release kinetics and dose-efficient sirolimus delivery from the stent and biodegradable platform contributes to early strut coverage and healing, as shown in the first-in-man OCT study. In future this may significantly shorten the duration of double antiplatelet therapy.

Acknowledgements

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Conflict of interest: Piotr P. Buszman — unrestricted research grants, Balton, Warsaw, Poland; Krzysztof Milewski — unrestricted research grants and consultancy, Balton, Warsaw, Poland.

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Zależna od dawki odpowiedź ściany naczyniowej na implantację stentu szybko uwalniającego lek, pokrytego polimerem biodegradowalnym ze zwiększonną dawką sirolimusu: badanie eksperymentalne na modelu restenozy tętnic wieńczych świń domowej

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Streszczenie

Wstęp: Chociaż stenty powlekane analogami rapamycyny o szybkim profilu uwalniania leku udowodniły swoje bezpieczeństwo, to ich skuteczność w zapobieganiu restenozie jest niewystarczająca.

Cel: Celem pracy była ocena efektów tkankowych w odpowiedzi na implantację stentu szybko uwalniającego lek, pokrytego polimerem biodegradowalnym ze zwiększonną dawką sirolimusu (ed-frSES, Alex, Balton) z wykorzystaniem modelu restenozy tętnic wieńczych świń domowej.

Metody: Wszczepiono 19 stentów ze 120% przerozmiarowaniem (tzw. overstretch) do tętnic wieńczych 7 świń domowych: 7 stentów typu ed-frSES z 1,3 μg/mm² sirolimusu, 8 stentów szybko uwalniających lek ze standardową dawką sirolimusu (frSES) i 8 stentów metalowych (BMS). Po 28 dniach obserwacji wykonano angiografię z oceną ilościową. Następnie zwierzęta zostały poddane eutanazji, a badane segmenty wypreparowane w celu przeprowadzenia analizy histopatologicznej.

 Wyniki: Po 28 dniach późna utrata światła w ocenie angiograficznej była najniższa w grupie ze zwiększoną dawką sirolimusu (ed-frSES: 0,20 ± 0,2 vs. frSES: 0,80 ± 0,5 vs. BMS: 0,96 ± 0,5 mm; p < 0,01). Podobne wyniki uzyskano w analizie histopatologicznej, w której obserwowano istotnie mniejsze procentowe zwiększenie pola przekroju w grupie ed-frSES (ed-frSES: 22,4 ± 12,7% vs. frSES: 35 ± 10,7% vs. BMS: 47,5 ± 12,5%; p < 0,01). W żadnej z badanych grup nie zaobserwowano okołoprzesłowego nacieku zapalnego. Poziom endotelializacji był nieco mniejszy w grupie ed-frSES, jednak różnica ta nie była istotna statystycznie (ed-frSES: 2,93 vs. frSES: 3 vs. BMS: 3; p = 0,05). Ponadto w grupie ed-frSES zaobserwowano okołoprzesłowo nieznaczną ilość włóknika (ed-frSES: 0,4 vs. frSES: 0 vs. BMS: 0; p = 0,05).

Wnioski: Potwierdzono zależność od dawki sirolimusu odpowiedź tkankową. Stenty szybko uwalniające lek ze zwiększonną dawką sirolimusu (ed-frSES) umożliwiają zadowalające gojenie się ściany naczyniowej, podobne do stentów ze standardową dawką leku, przy jednocześnie skuteczniejszym hamowaniu restenozy.

Słowa kluczowe: stenty uwalniające sirolimus, szybkie uwalnianie, polimer biodegradowalny, model tętnic wieńczych świń

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