SHORT COMMUNICATION

Association of cardiac troponin I with prothrombotic alterations in atrial fibrillation

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INTRODUCTION

Atrial fibrillation (AF) increases the risk of stroke and systemic thromboembolism. A hypercoagulable state in AF is reflected by elevated von Willebrand factor (vWF), D-dimer, and thrombin generation (TG), as well as increased platelet activation [1]. The usefulness of several biomarkers in stroke and bleeding risk prediction among AF patients, in particular N-terminal pro–B-type natriuretic peptide (NT-proBNP) and high-sensitivity cardiac troponin I (cTnI-hs), has been demonstrated, although practical implications of this strategy are uncertain [2].

We sought to assess the relations of four such markers: cTnI-hs, cystatin C, vWF, and NT-proBNP, with prothrombotic alterations in AF patients.

METHODS

We investigated 65 patients with AF free of myocardial infarction or venous thromboembolism within the previous three months, with creatinine clearance ≥ 15 mL/min, and without liver cirrhosis, known cancer, or acute infection. Detailed methodology of the study was presented previously [3]. Vitamin K antagonists and low-molecular-weight heparin (the last dose ≥ 12 h) were discontinued before the blood draw. The study was approved by the Bioethical Committee, and patients gave informed consent.

Venous blood samples were taken with minimal stasis, and routine blood tests were performed by standard laboratory techniques. Electrochemiluminescence immunoassay (Roche Diagnostics, Mannheim, Germany) was used to measure NT-proBNP levels. The ARCHITECT i1000SR and ARCHITECT ci8200 (Abbott Laboratories, Abbott Park, IL, USA) were used to assess cTnI-hs and cystatin C levels, respectively. Latex immunoassay using a STAR coagulation instrument (Diagnostica Stago, Asnières, France) was used to measure vWF antigen.

Assessment of TG profile was performed as described [4]. Briefly, corn trypsin inhibitor was added to citrated plasma (a final concentration of 0.1 mg/mL), and then samples (80 µL each) were mixed with relipidated tissue factor to achieve a final concentration of 5 pM and with a 2.5 mM Z-GGR-AMC/90 mM CaCl₂ solution in Hepes-buffered saline to achieve a 417 µM and 15 mM final concentration, respectively, followed by incubation at 37°C for 3 min. Lastly, a phospholipid vesicles solution in Hepes-buffered saline was added to samples to achieve 20 µM final concentration, thereby initiating TG. Hydrolysis of the 7-amino-4-methylcoumarin substrate at 370 nm excitation and 460 nm emission wavelengths was followed for 3600 s. A calibration curve built by sequential dilutions of human thrombin into pooled multi-donor plasma from healthy individuals was used to convert substrate hydrolysis to thrombin concentration. We used the BioTek Synergy 4 plate reader and Gen5 plate reader software for analysis (BioTek, Winooski, VT, USA).

Statistical analysis

Quantitative variables were presented as the mean (standard deviation) or median (interquartile range). Normal distribution was assessed using the Shapiro-Wilk test. Quantitative variables were compared using Student t-test or Mann-Whitney U test, and correlations were tested using a linear Pearson correlation or Spearman’s rank test, as appropriate. We divided patients into three groups based on tertiles of cTnI-hs levels, and into four groups based on the CHA2DS2-VASc score. Normally distributed data in three or more groups

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were compared using one-way ANOVA (in homogeneity of variance) or Welch ANOVA (otherwise). Tukey’s HSD test was used to assess pairwise comparisons. For comparison of non-normally distributed variables in three or more groups, the Kruskal-Wallis test was used, and for pairwise comparisons the Kruskal-Wallis test with the Bonferroni correction was used. Qualitative variables were described by number (frequency) and compared by χ² test or Fisher exact test. To assess predictors of peak thrombin concentration we performed linear regression analysis. R² was calculated, and assessment of model adequacy using F test was performed. P values of < 0.05 were considered statistically significant. IBM SPSS Statistics (version 24.0, IBM Corp., Armonk, NY, USA) was used to perform statistical analysis.

RESULTS AND DISCUSSION
The current study group included 65 patients aged 69.8 ± 9.9 years (41.5% women), who had mostly permanent AF (53.8%) (Suppl. Table 1 — see journal website). CHA2DS2-VASc score ≥ 2 was observed in 93.8% of patients.

Patients in the third cTnI-hs tertile were older than those in the first and second tertiles (all p < 0.005). Patients in the first cTnI-hs tertile had a higher haemoglobin concentration than those in the third cTnI-hs tertile (p = 0.02) (Suppl. Table 2 — see journal website). In the whole group cystatin C was correlated with cTnI-hs (r = 0.34, p = 0.005), but not with NT-proBNP (r = 0.12, p = 0.33).

CHA2DS2-VASc score was positively correlated with cTnI-hs (R = 0.47, p < 0.001) and cystatin C (r = 0.39,
p = 0.001), with a similar trend for NT-proBNP (R = 0.22, p = 0.07). cTnl-hs and cystatin C (Fig. 1A, B) were higher in subjects with the highest CHA2DS2-VASc score when compared to patients with the lowest CHA2DS2-VASc score. No such associations were observed for NT-proBNP, vWF, creatinine, or TG parameters. There was no correlation between vWF and CHA2DS2-VASc score, cTnl-hs, cystatin C, or NT-proBNP (data not shown).

Analysis of TG showed that lag phase and endogenous thrombin potential did not correlate with cTnl-hs, NT-proBNP, or cystatin C. Time to peak thrombin generation was correlated with vWF (Fig. 1C) and tended to correlate with cTnl-hs (r = −0.22, p = 0.08), but not with NT-proBNP or cystatin C. Importantly, peak thrombin concentration correlated with cTnl-hs and was associated with cTnl-hs tertiles (R = 0.37, p = 0.003, Fig. 1D) but not with NT-proBNP, cystatin C, or vWF. Peak thrombin concentration was not related to glomerular filtration rate (GFR) and age (r = 0.04, p = 0.73 and r = 0.10, p = 0.44, respectively). In linear regression analysis peak thrombin concentration was predicted by cTnl-hs (unstandardized coefficient B: 17.09; 95% confidence interval [CI]: 0.06–34.11, p = 0.049) but not GFR (B: 0.43; 95% CI: −2.02–2.88; p = 0.73) or age (B: 1.84; 95% CI: −2.86–6.54; p = 0.44). Peak thrombin concentration was predicted by cTnl-hs (B: 27.30; 95% CI: 7.15–47.45; p = 0.009), after inclusion of GFR in the model (B: 2.55; 95% CI: −0.26–5.37; p = 0.074), R² = 0.11, F = 3.73, p = 0.029.

Our study shows that peak thrombin concentration is associated with cTnl-hs in AF patients, suggesting that the most sensitive TG parameter is associated with processes leading to myocardial injury, strain, or ischaemia in this common arrhythmia. This sensitivity of peak thrombin, but not endogenous thrombin potential, is best reflected by the observation that in patients with acute coronary syndromes at admission and six months later the sole elevated TG marker is peak thrombin when compared to controls [5]. Peak thrombin generated, together with shorter times to maximum levels, are well-established features of hypercoagulable states and may be useful in the assessment of bleeding and thromboembolic risk [6]. Our preliminary study suggests that cardiac troponins better reflect TG in AF compared with other biomarkers. In AF elevation of cTnl-hs is related to cardiac disease, including heart failure and coronary artery disease, the risk factors for stroke in AF, being associated with the severity of myocardial ischaemia and/or injury [7, 8]. Troponin release may also occur during tachyarrhythmias and bradyarrhythmias frequently observed in patients with AF [9].

We confirmed that cTnl-hs and renal function parameters are associated with CHA2DS2-VASc score [10, 11] and that higher AF burden is associated with elevated troponin levels [7].

Our study supports biomarker-based risk stratification among patients with AF [2, 12]. Current results and our previous observations indicate that cardiac troponin and NT-proBNP may be implicated in similar pathogenic pathways leading to faster and/or enhanced TG, which significantly contributes to a hypercoagulable state in AF [3]. These two biomarkers and associated pathways may be common with those noted in ischaemic stroke [13].

The current study has several limitations. First, the study group was small. Secondly, TG can be determined using various assays, including the use of different concentrations of reagents in calibrated automated thrombograms. We did not measure anti-Xa activity in our study. It is unclear whether anticoagulated AF patients show similar associations between TG variables and other biomarkers.

The current study provides insights into relations of circulating biomarkers with thromboembolic risk and thrombin generation in patients with AF.

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7. Hijazi Z, Siegbahn A, Andersson U, et al. Comparison of reagents in calibrated automated thrombograms. We did not measure anti-Xa activity in our study. It is unclear whether anticoagulated AF patients show similar associations between TG variables and other biomarkers.


